



19th International Conference — Science, Technology and Innovation *Booklets*



RENIACYT - LATINDEX - Research Gate - DULCINEA - CLASE - Sudoc - HISPANA - SHERPA UNIVERSIA - Google Scholar DOI - REDIB - Mendeley - DIALNET - ROAD - ORCID

Title: BDNF expression in blood. Study in iron deficient females

Authors: VIEYRA-REYES, Patricia and GÓMEZ-LAGUNAS, Néstor G.

Editorial label ECORFAN: 607-8695

BECORFAN Control Number: 2022-01

BECORFAN Classification (2022): 131222-0001

Pages: 10

RNA: 03-2010-032610115700-14

ECORFAN-México, S.C.

143 – 50 Itzopan Street

La Florida, Ecatepec Municipality

Mexico State, 55120 Zipcode

Phone: +52 1 55 6159 2296

Skype: ecorfan-mexico.s.c.

E-mail: contacto@ecorfan.org

Facebook: ECORFAN-México S. C.

Twitter: @EcorfanC

www.ecorfan.org

Holdings

Mexico	Colombia	Guatemala
Bolivia	Cameroon	Democratic
Spain	El Salvador	Republic
Ecuador	Taiwan	of Congo
Peru	Paraguay	Nicaragua

Introducción

- 🧠 La deficiencia de hierro (DFe) es la deficiencia de micronutrientes más común en todo el mundo.
- 🧠 Afecta a 614 millones de mujeres y 280 millones de niños (WHO, 2020a).
- 🧠 Diversos trastornos neurocognitivos relacionados con deficiencia de hierro, están vinculados a alteraciones en factores neurotróficos.
- 🧠 El factor neurotrófico derivado del cerebro, BDNF, es un polipéptido que actúa como factor de crecimiento modulando la síntesis, metabolismo y liberación de neurotransmisores (Crump et al., 2014), supervivencia, diferenciación y plasticidad sináptica en los sistemas nerviosos central y periférico (Lu & Figurov, 1997; Morse et al., 2015; Texel et al., 2011).

- 🧠 Se ha demostrado una baja expresión y actividad a largo plazo del BDNF a nivel de hipocampo en sujetos con DFe (Tran et al., 2009).
- 🧠 BDNF puede atravesar la barrera hematoencefálica; por lo tanto, los niveles en suero y en líquido cefalorraquídeo están fuertemente correlacionados con los niveles cerebrales (Gururajan et al., 2014; Harris & Barraclough, 1997; St Laurent et al., 2013).
- 🧠 El gen de BDNF está presente un elemento de respuesta al estrógeno (ERE), por lo que los estrógenos aumentan la expresión de este factor neurotrófico (Singh et al., 1995; Sohrabji et al., 1995).
- 🧠 Pocos estudios se realizan en hembras, con respecto a deficiencia de hierro se desconoce si en éstas la deficiencia crónica de este elemento traza afecta sobre los niveles de BDNF en sangre, un determinante indirecto de los niveles en cerebro.

Metodología

Pie de cría

Grupo Deficiente de hierro (Dfe): 20 ratas hembras (de 3 meses de edad o 250 g) alimentadas con dieta deficiente en hierro (10 ppm FeSO₄, Lab Diets AIN-76W / 10) 14 días antes del apareamiento.

Grupo control: 10 ratas hembras alimentadas con dieta control (100 ppm FeSO₄, Lab diets AIN-76W / 100) 14 días antes del apareamiento.

Crías

21 días después del nacimiento (DPN), las crías fueron destetadas. Las hembras fueron seleccionadas para el presente experimento, los machos fueron empleados en otros proyectos. Las crías hembras se mantuvieron con el mismo tipo de dieta ofrecida a sus madres hasta los 70 DPN; con la excepción del grupo suplementado “DFe+S”, un conjunto de crías hembras ID, que recibieron del 21 al 70 DPN dieta control.

Grupo control



Grupo DFe



Grupo Dfe+S



70 PND

Eutanasia

Obtención de muestra
sanguínea

**Determinación de Hierro
unido a hemoglobina (Fe-Hb)**

Determinación de BDNF en sangre

Resultados

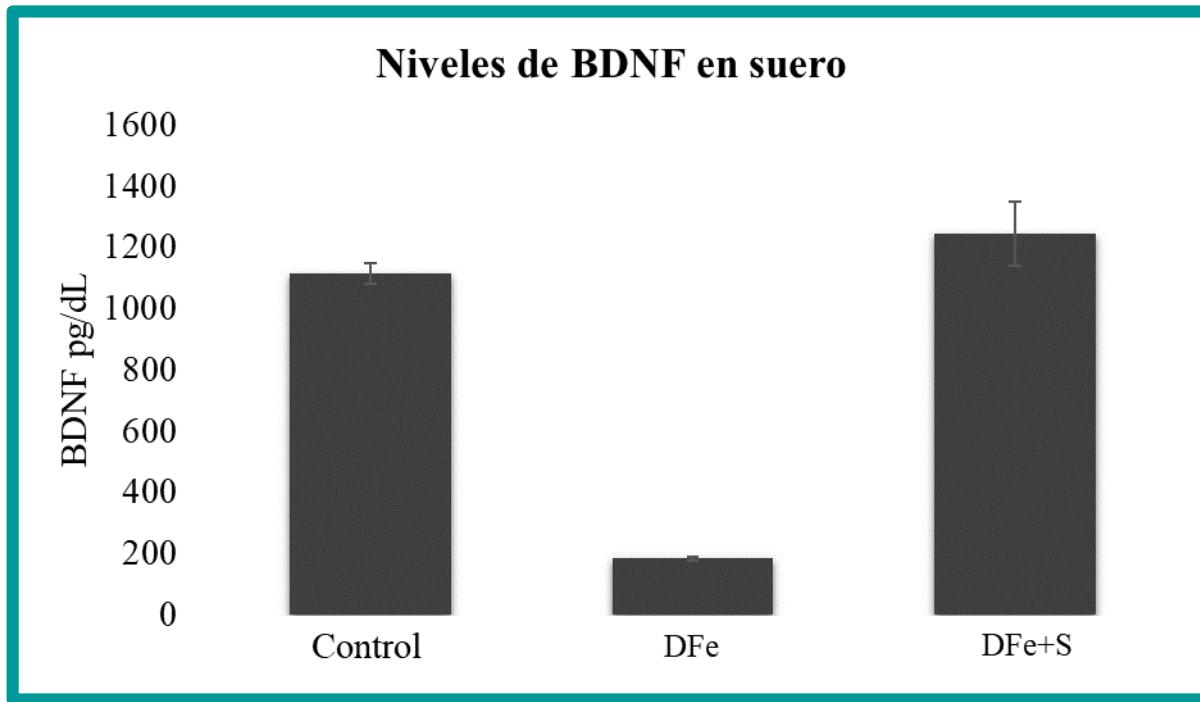
Determinación de hierro unido a hemoglobina

Al estudiar al grupo ID con respecto al grupo control, se encontró que los primeros presentan 10.9% menos de Fe-Hb y 3.8% menos que el grupo ID+S, ver Tabla 1.

Grupo	Fe - Hb (mg/kg PV)
Control	3.71±0.11
Deficiente de hierro+suplemento	3.47±0.11
Deficiente de hierro	3.18±0.23*

Tabla 1. Niveles de hierro unido a hemoglobina “Fe-Hb”

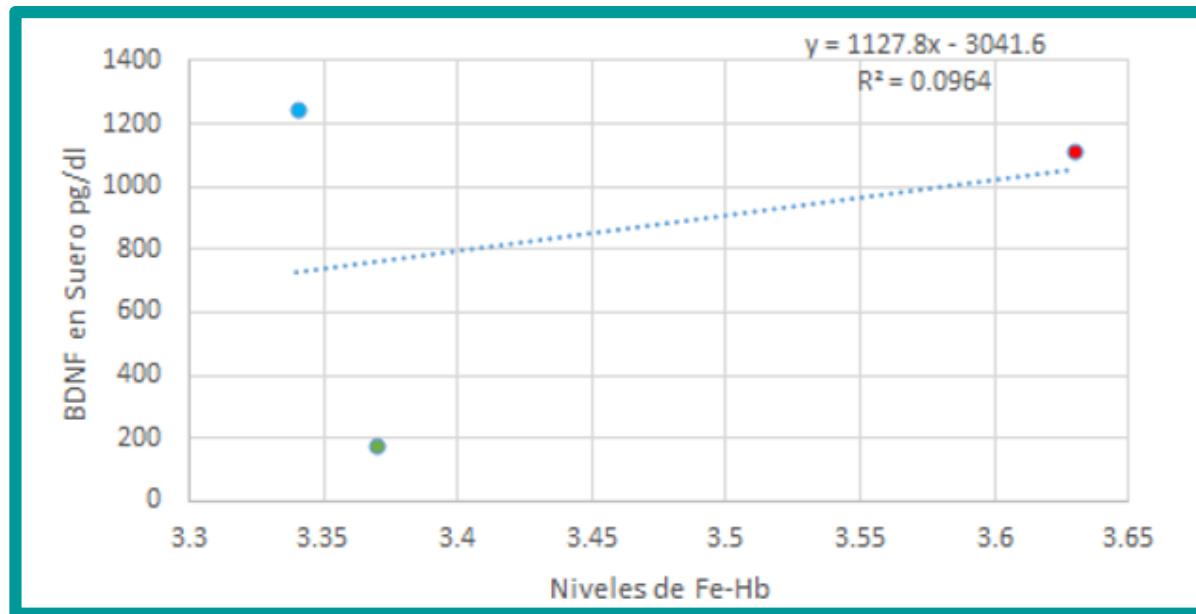
* vs. Hembra del grupo control ($p \leq 0.05$)



Gráfica 1. Niveles de BDNF en sangre. Los valores indican la media \pm error estándar.

* vs. Hembra del grupo control ($p \leq 0.05$)

**vs. Hembra del grupo Deficiente de hierro+Suplemento ($p \leq 0.05$)



Gráfica 2. Correlación entre los niveles de BDNF y los de hierro unido a hemoglobina. Los puntos en rojo indican hembras control; verdes DFe y azules DFe+S.

Conclusiones

- A. Ante una deficiencia de hierro crónica, los niveles de BDNF disminuyen periféricamente.
- B. La suplementación con hierro en sujetos con deficiencia normaliza los niveles de BDNF a nivel periférico.
- C. Los niveles de hierro unido a hemoglobina se correlacionan positivamente con los niveles de BDNF en sangre.

Referencias

- Araya, A. V., Orellana, X., & Espinoza, J. (2008). Evaluation of the effect of caloric restriction on serum BDNF in overweight and obese subjects: preliminary evidences. *Endocrine*, 33(3), 300-304. <https://doi.org/10.1007/s12020-008-9090-x>
- Barks, A., Fretham, S. J. B., Georgieff, M. K., & Tran, P. V. (2018). Early-Life Neuronal-Specific Iron Deficiency Alters the Adult Mouse Hippocampal Transcriptome. *J Nutr*, 148(10), 1521-1528. <https://doi.org/10.1093/jn/nxy125>
- Barks, A., Hall, A. M., Tran, P. V., & Georgieff, M. K. (2019). Iron as a model nutrient for understanding the nutritional origins of neuropsychiatric disease. *Pediatr Res*, 85(2), 176-182. <https://doi.org/10.1038/s41390-018-0204-8>
- Blegen, M. B., Kennedy, B. C., Thibert, K. A., Gewirtz, J. C., Tran, P. V., & Georgieff, M. K. (2013). Multigenerational effects of fetal-neonatal iron deficiency on hippocampal BDNF signaling. *Physiol Rep*, 1(5), e00096. <https://doi.org/10.1002/phy2.96>
- Brunette, K. E., Tran, P. V., Wobken, J. D., Carlson, E. S., & Georgieff, M. K. (2010). Gestational and neonatal iron deficiency alters apical dendrite structure of CA1 pyramidal neurons in adult rat hippocampus. *Dev Neurosci*, 32(3), 238-248. <https://doi.org/10.1159/000314341>
- Burden, M. J., Westerlund, A. J., Armony-Sivan, R., Nelson, C. A., Jacobson, S. W., Lozoff, B., Angelilli, M. L., & Jacobson, J. L. (2007). An event-related potential study of attention and recognition memory in infants with iron-deficiency anemia. *Pediatrics*, 120(2), e336-345. <https://doi.org/10.1542/peds.2006-2525>
- Carlson, E. S., Stead, J. D., Neal, C. R., Petryk, A., & Georgieff, M. K. (2007). Perinatal iron deficiency results in altered developmental expression of genes mediating energy metabolism and neuronal morphogenesis in hippocampus. *Hippocampus*, 17(8), 679-691. <https://doi.org/10.1002/hipo.20307>
- Carlson, E. S., Tkac, I., Magid, R., O'Connor, M. B., Andrews, N. C., Schallert, T., Gunshin, H., Georgieff, M. K., & Petryk, A. (2009). Iron is essential for neuron development and memory function in mouse hippocampus. *J Nutr*, 139(4), 672-679. <https://doi.org/10.3945/jn.108.096354>
- Crump, C., Sundquist, K., Sundquist, J., & Winkleby, M. A. (2014). Sociodemographic, psychiatric and somatic risk factors for suicide: a Swedish national cohort study. *Psychol Med*, 44(2), 279-289. <https://doi.org/10.1017/s0033291713000810>
- Faux, N. G., Rembach, A., Wiley, J., Ellis, K. A., Ames, D., Fowler, C. J., Martins, R. N., Pertile, K. K., Rumble, R. L., Trounson, B., Masters, C. L., Group, A. R., & Bush, A. I. (2014). An anemia of Alzheimer's disease. *Mol Psychiatry*, 19(11), 1227-1234. <https://doi.org/10.1038/mp.2013.178>
- Fretham, S. J., Carlson, E. S., & Georgieff, M. K. (2011). The role of iron in learning and memory. *Adv Nutr*, 2(2), 112-121. <https://doi.org/10.3945/an.110.000190>
- Fretham, S. J., Carlson, E. S., Wobken, J., Tran, P. V., Petryk, A., & Georgieff, M. K. (2012). Temporal manipulation of transferrin-receptor-1-dependent iron uptake identifies a sensitive period in mouse hippocampal neurodevelopment. *Hippocampus*, 22(8), 1691-1702. <https://doi.org/10.1002/hipo.22004>
- Harris, E. C., & Barraclough, B. (1997). Suicide as an outcome for mental disorders. A meta-analysis. *Br J Psychiatry*, 170, 205-228.
- Hernandez, M., Sousa, V., Villalpando, S., Moreno, A., Montalvo, I., & Lopez-Alarcon, M. (2006). Cooking and Fe fortification have different effects on Fe bioavailability of bread and tortillas. *J Am Coll Nutr*, 25(1), 20-25. [https://doi.org/25/1/20 \[pii\]](https://doi.org/25/1/20 [pii])
- Insel, B. J., Schaefer, C. A., McKeague, I. W., Susser, E. S., & Brown, A. S. (2008). Maternal iron deficiency and the risk of schizophrenia in offspring. *Arch Gen Psychiatry*, 65(10), 1136-1144. <https://doi.org/10.1001/archpsyc.65.10.1136>
- Kuzawa, C. W. (1998). Adipose tissue in human infancy and childhood: an evolutionary perspective. *Am J Phys Anthropol*, Suppl 27, 177-209. [https://doi.org/10.1002/\(sici\)1096-8644\(1998\)107:27+<177::aid-ajpa7>3.0.co;2-b](https://doi.org/10.1002/(sici)1096-8644(1998)107:27+<177::aid-ajpa7>3.0.co;2-b)
- Lee, J., Duan, W., & Mattson, M. P. (2002). Evidence that brain-derived neurotrophic factor is required for basal neurogenesis and mediates, in part, the enhancement of neurogenesis by dietary restriction in the hippocampus of adult mice. *J Neurochem*, 82(6), 1367-1375.
- Lee, M. Y., Lee, J., Hyeon, S. J., Cho, H., Hwang, Y. J., Shin, J. Y., McKee, A. C., Kowall, N. W., Kim, J. I., Stein, T. D., Hwang, D., & Ryu, H. (2020). Epigenome signatures landscaped by histone H3K9me3 are associated with the synaptic dysfunction in Alzheimer's disease. *Aging Cell*, 19(6), e13153. <https://doi.org/10.1111/acel.13153>
- Li, Z., Okamoto, K., Hayashi, Y., & Sheng, M. (2004). The importance of dendritic mitochondria in the morphogenesis and plasticity of spines and synapses. *Cell*, 119(6), 873-887. <https://doi.org/10.1016/j.cell.2004.11.003>
- Liu, S. X., Barks, A. K., Lunos, S., Gewirtz, J. C., Georgieff, M. K., & Tran, P. V. (2021). Prenatal Iron Deficiency and Choline Supplementation Interact to Epigenetically Regulate Jarid1b and Bdnf in the Rat Hippocampus into Adulthood. *Nutrients*, 13(12). <https://doi.org/10.3390/nu13124527>
- Lozoff, B. (2011). Early iron deficiency has brain and behavior effects consistent with dopaminergic dysfunction. *J Nutr*, 141(4), 740s-746s. <https://doi.org/10.3945/jn.110.131169>
- Lozoff, B., & Georgieff, M. K. (2006). Iron deficiency and brain development. *Semin Pediatr Neurol*, 13(3), 158-165. <https://doi.org/10.1016/j.spen.2006.08.004>
- Lu, B., & Figurov, A. (1997). Role of neurotrophins in synapse development and plasticity. *Rev Neurosci*, 8(1), 1-12. <http://www.ncbi.nlm.nih.gov/pubmed/9402641>
- McAllister, A. K., Katz, L. C., & Lo, D. C. (1999). Neurotrophins and synaptic plasticity. *Annu Rev Neurosci*, 22, 295-318. <https://doi.org/10.1146/annurev.neuro.22.1.295>
- Morse, S. J., Butler, A. A., Davis, R. L., Soller, I. J., & Lubin, F. D. (2015). Environmental enrichment reverses histone methylation changes in the aged hippocampus and restores age-related memory deficits. *Biology (Basel)*, 4(2), 298-313. <https://doi.org/10.3390/biology4020298>

- Riggins, T., Miller, N. C., Bauer, P. J., Georgieff, M. K., & Nelson, C. A. (2009). Consequences of low neonatal iron status due to maternal diabetes mellitus on explicit memory performance in childhood. *Dev Neuropsychol*, 34(6), 762-779.
<https://doi.org/10.1080/87565640903265145>
- Scharfman, H. E., & MacLusky, N. J. (2006). Estrogen and brain-derived neurotrophic factor (BDNF) in hippocampus: complexity of steroid hormone-growth factor interactions in the adult CNS. *Front Neuroendocrinol*, 27(4), 415-435.
<https://doi.org/10.1016/j.yfrne.2006.09.004>
- Schmidt, A. T., Alvarez, G. C., Grove, W. M., Rao, R., & Georgieff, M. K. (2012). Early iron deficiency enhances stimulus-response learning of adult rats in the context of competing spatial information. *Dev Cogn Neurosci*, 2(1), 174-180.
<https://doi.org/10.1016/j.dcn.2011.07.014>
- Schmidt, R. J., Tancredi, D. J., Krakowiak, P., Hansen, R. L., & Ozonoff, S. (2014). Maternal intake of supplemental iron and risk of autism spectrum disorder. *Am J Epidemiol*, 180(9), 890-900. <https://doi.org/10.1093/aje/kwu208>
- Singh, M., Meyer, E. M., & Simpkins, J. W. (1995). The effect of ovariectomy and estradiol replacement on brain-derived neurotrophic factor messenger ribonucleic acid expression in cortical and hippocampal
- Sohrabji, F., Miranda, R. C., & Toran-Allerand, C. D. (1995). Identification of a putative estrogen response element in the gene encoding brain-derived neurotrophic factor. *Proc Natl Acad Sci U S A*, 92(24), 11110-11114.
<http://www.ncbi.nlm.nih.gov/pubmed/7479947>
- Solati, Z., Jazayeri, S., Tehrani-Doost, M., Mahmoodianfarid, S., & Gohari, M. R. (2015). Zinc monotherapy increases serum brain-derived neurotrophic factor (BDNF) levels and decreases depressive symptoms in overweight or obese subjects: a double-blind, randomized, placebo-controlled trial. *Nutr Neurosci*, 18(4), 162-168. <https://doi.org/10.1179/1476830513y.0000000105>
- St Laurent, R., Helm, S. R., & Glenn, M. J. (2013). Reduced cocaine-seeking behavior in heterozygous BDNF knockout rats. *Neurosci Lett*, 544, 94-99. <https://doi.org/10.1016/j.neulet.2013.03.050>
- Texel, S. J., Zhang, J., Camandola, S., Unger, E. L., Taub, D. D., Koehler, R. C., Harris, Z. L., & Mattson, M. P. (2011). Ceruloplasmin deficiency reduces levels of iron and BDNF in the cortex and striatum of young mice and increases their vulnerability to stroke. *PLoS One*, 6(9), e25077. <https://doi.org/10.1371/journal.pone.0025077>
- Tran, P. V., Carlson, E. S., Freetham, S. J. B., & Georgieff, M. K. (2008). Early-Life Iron Deficiency Anemia Alters Neurotrophic Factor Expression and Hippocampal Neuron Differentiation in Male Rats. *The Journal of Nutrition*, 138(12), 2495-2501.
<https://doi.org/10.3945/jn.108.091553>
- Tran, P. V., Freetham, S. J., Carlson, E. S., & Georgieff, M. K. (2009). Long-term reduction of hippocampal brain-derived neurotrophic factor activity after fetal-neonatal iron deficiency in adult rats. *Pediatr Res*, 65(5), 493-498.
<https://doi.org/10.1203/PDR.0b013e31819d90a1>
- Tran, P. V., Kennedy, B. C., Lien, Y. C., Simmons, R. A., & Georgieff, M. K. (2015). Fetal iron deficiency induces chromatin remodeling at the Bdnf locus in adult rat hippocampus. *Am J Physiol Regul Integr Comp Physiol*, 308(4), R276-282.
<https://doi.org/10.1152/ajpregu.00429.2014>
- Tran, P. V., Kennedy, B. C., Pisansky, M. T., Won, K. J., Gewirtz, J. C., Simmons, R. A., & Georgieff, M. K. (2016). Prenatal Choline Supplementation Diminishes Early-Life Iron Deficiency-Induced Reprogramming of Molecular Networks Associated with Behavioral Abnormalities in the Adult Rat Hippocampus. *J Nutr*, 146(3), 484-493.
<https://doi.org/10.3945/jn.115.227561>
- Unger, E. L., Paul, T., Murray-Kolb, L. E., Felt, B., Jones, B. C., & Beard, J. L. (2007). Early iron deficiency alters sensorimotor development and brain monoamines in rats. *J Nutr*, 137(1), 118-124. [https://doi.org/137/1/118\[pii\]](https://doi.org/137/1/118[pii])
- WHO. (2020a). WHO guidance helps detect iron deficiency and protect brain development.
<https://www.who.int/news-room/20-04-2020-who-guidance-helps-detect-iron-deficiency-and-protect-brain-development>
- WHO. (2020b). WHO guideline on use of ferritin concentrations to assess iron status in individuals and populations.
<https://www.who.int/publications/item/9789240000124>
- Wienk, K. J., Marx, J. J., & Beynen, A. C. (1999). The concept of iron bioavailability and its assessment. *Eur J Nutr*, 38(2), 51-75. http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&dopt=Citation&list_uids=10352945
- Zhang, H. Y., Song, N., Jiang, H., Bi, M. X., & Xie, J. X. (2014). Brain-derived neurotrophic factor and glial cell line-derived neurotrophic factor inhibit ferrous iron influx via divalent metal transporter 1 and iron regulatory protein 1 regulation in ventral mesencephalic neurons. *Biochim Biophys Acta*, 1843(12), 2967-2975. <https://doi.org/10.1016/j.bbamcr.2014.09.010>



ECORFAN®

© ECORFAN-Mexico, S.C.

No part of this document covered by the Federal Copyright Law may be reproduced, transmitted or used in any form or medium, whether graphic, electronic or mechanical, including but not limited to the following: Citations in articles and comments Bibliographical, compilation of radio or electronic journalistic data. For the effects of articles 13, 162,163 fraction I, 164 fraction I, 168, 169,209 fraction III and other relative of the Federal Law of Copyright. Violations: Be forced to prosecute under Mexican copyright law. The use of general descriptive names, registered names, trademarks, in this publication do not imply, uniformly in the absence of a specific statement, that such names are exempt from the relevant protector in laws and regulations of Mexico and therefore free for General use of the international scientific community. BECORFAN is part of the media of ECORFAN-Mexico, S.C., E: 94-443.F: 008- (www.ecorfan.org/booklets)